# The biomaRt user's guide

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# 1 Introduction

In recent years a wealth of biological data has become available in public data repositories. Easy access to these valuable data resources and firm integration with data analysis is needed for comprehensive bioinformatics data analysis. The *biomaRt* package, provides an interface to a growing collection of databases implementing the BioMart software suite (`http://www.biomart.org`). The package enables retrieval of large amounts of data

in a uniform way without the need to know the underlying database schemas or write complex SQL queries. Examples of BioMart databases are Ensembl, Uniprot and HapMap. These major databases give biomaRt users direct access to a diverse set of data and enable a wide range of powerful online queries from R.

## 2 Selecting a BioMart database and dataset

Every analysis with *biomaRt* starts with selecting a BioMart database to use. A first step is to check which BioMart web services are available. The function `listMarts` will display all available BioMart web services

```
> library("biomaRt")
> listMarts()
```

| | biomart | version |
|---|---|---|
| 1 | ensembl | ENSEMBL GENES 62 (SANGER UK) |
| 2 | snp | ENSEMBL VARIATION 62 (SANGER UK) |
| 3 | functional_genomics | ENSEMBL FUNCTIONAL GENOMICS 62 (SANGER UK) |
| 4 | vega | VEGA 42 (SANGER UK) |
| 5 | bacterial_mart_9 | ENSEMBL BACTERIA 9 (EBI UK) |
| 6 | fungal_mart_9 | ENSEMBL FUNGAL 9 (EBI UK) |
| 7 | metazoa_mart_9 | ENSEMBL METAZOA 9 (EBI UK) |
| 8 | plant_mart_9 | ENSEMBL PLANT 9 (EBI UK) |
| 9 | protist_mart_9 | ENSEMBL PROTISTS 9 (EBI UK) |
| 10 | msd | MSD (EBI UK) |
| 11 | htgt | WTSI MOUSE GENETICS PROJECT (SANGER UK) |
| 12 | REACTOME | REACTOME (CSHL US) |
| 13 | WS220-testing | WORMBASE 220 (CSHL US) |
| 14 | dicty | DICTYBASE (NORTHWESTERN US) |
| 15 | biomart | MGI (JACKSON LABORATORY US) |
| 16 | g4public | HGNC (EBI UK) |
| 17 | pride | PRIDE (EBI UK) |
| 18 | prod-intermart_2 | INTERPRO (EBI UK) |
| 19 | unimart | UNIPROT (EBI UK) |
| 20 | biomartDB | PARAMECIUM GENOME (CNRS FRANCE) |
| 21 | Eurexpress Biomart | EUREXPRESS (MRC EDINBURGH UK) |
| 22 | pepseekerGOLD_mart06 | PEPSEEKER (UNIVERSITY OF MANCHESTER UK) |
| 23 | Potato_01 | DB POTATO (INTERNATIONAL POTATO CENTER-CIP) |
| 24 | Sweetpotato_01 | DB SWEETPOTATO (INTERNATIONAL POTATO CENTER-CIP) |
| 25 | phytozome_mart | PHYTOZOME (JGI/CIG US) |
| 26 | cyanobase_1 | CYANOBASE 1 (KAZUSA JAPAN) |
| 27 | rhizobase_2 | RHIZOBASE 2 (KAZUSA) |
| 28 | cyanobase_2_togo | CYANOBASE 2 TOGO (KAZUSA) |
| 29 | rhizobase_2_togo | RHIZOBASE 2 TOGO (KAZUSA) |
| 30 | plant_genome_1 | PLANT GENOME 1 (KAZUSA) |
| 31 | plant_genome_1_togo | PLANT GENOMES 1 TOGO (KAZUSA) |
| 32 | HapMap_rel27 | HAPMAP 27 (NCBI US) |
| 33 | CosmicMart | COSMIC (SANGER UK) |
| 34 | cildb_all_v2 | CILDB INPARANOID AND FILTERED BEST HIT (CNRS FRANCE) |
| 35 | cildb_inp_v2 | CILDB INPARANOID (CNRS FRANCE) |

```
36                          experiments                                      INTOGEN EXPERIMENTS
37                         combinations                                    INTOGEN COMBINATIONS
38                          gmap_japonica                RICE-MAP JAPONICA (PEKING UNIVESITY CHINA)
39                 europhenomeannotations                                           EUROPHENOME
40                          emma_biomart                  THE EUROPEAN MOUSE MUTANT ARCHIVE (EMMA)
41                                 ikmc                          IKMC GENES AND PRODUCTS (IKMC)
42                  EMAGE gene expression                                   EMAGE GENE EXPRESSION
43                  EMAP anatomy ontology                                   EMAP ANATOMY ONTOLOGY
44                EMAGE browse repository                                 EMAGE BROWSE REPOSITORY
45                              GermOnline                                               GERMONLINE
46                      vectorbase_mart_60                                  VECTORBASE GENES 7
47                   vectorbase_snp_mart_60                               VECTORBASE VARIATION 7
48                              expression                              VECTORBASE EXPRESSION MART
49 Sigenae Oligo Annotation (Ensembl 59)                    SIGENGE OLIGO ANNOTATION (ENSEMBL 59)
50 Sigenae Oligo Annotation (Ensembl 56)                    SIGENGE OLIGO ANNOTATION (ENSEMBL 56)
51                              gmap_indica                 RICE-MAP INDICA (PEKING UNIVERSITY CHINA)
52                                Ensembl56 PANCREATIC EXPRESSION DATABASE (INSTITUTE OF CANCER  UK)
53                      ENSEMBL_MART_PLANT                GRAMENE 30 ENSEMBL GENES (CSHL/CORNELL US)
54                  ENSEMBL_MART_PLANT_SNP                    GRAMENE 30 VARIATION (CSHL/CORNELL US)
55                       GRAMENE_MARKER_30                      GRAMENE 30 MARKERS (CSHL/CORNELL US)
56                          GRAMENE_MAP_30                     GRAMENE 30 MAPPINGS (CSHL/CORNELL US)
57                                QTL_MART                       GRAMENE 32 QTL DB (CSHL/CORNELL US)
58                       salmosalar2_mart                  UNIGENE SALMO SALAR DATABASE (CMM CHILE)
59                            trucha_mart          UNIGENE ONCORHYNCHUS MYKISS DATABASE (CMM CHILE)
```

Note: if the function `useMart` runs into proxy problems you should set your proxy first before calling any biomaRt functions. You can do this using the Sys.putenv command:

```
Sys.putenv("http\_proxy" = "http://my.proxy.org:9999")
```

The `useMart` function can now be used to connect to a specified BioMart database, this must be a valid name given by `listMarts`. In the next example we choose to query the Ensembl BioMart database.

```
> ensembl = useMart("ensembl")
```

BioMart databases can contain several datasets, for Ensembl every species is a different dataset. In a next step we look at which datasets are available in the selected BioMart by using the function `listDatasets`.

```
> listDatasets(ensembl)
```

```
                       dataset                                 description      version
1         oanatinus_gene_ensembl       Ornithorhynchus anatinus genes (OANA5)        OANA5
2          tguttata_gene_ensembl    Taeniopygia guttata genes (taeGut3.2.4)  taeGut3.2.4
3        cporcellus_gene_ensembl            Cavia porcellus genes (cavPor3)      cavPor3
4        gaculeatus_gene_ensembl     Gasterosteus aculeatus genes (BROADS1)      BROADS1
5         lafricana_gene_ensembl         Loxodonta africana genes (loxAfr3)      loxAfr3
6        mlucifugus_gene_ensembl           Myotis lucifugus genes (myoLuc1)      myoLuc1
7          hsapiens_gene_ensembl             Homo sapiens genes (GRCh37.p3)    GRCh37.p3
```

```
8         choffmanni_gene_ensembl           Choloepus hoffmanni genes (choHof1)      choHof1
9          csavignyi_gene_ensembl                Ciona savignyi genes (CSAV2.0)      CSAV2.0
10            fcatus_gene_ensembl                       Felis catus genes (CAT)          CAT
11       rnorvegicus_gene_ensembl             Rattus norvegicus genes (RGSC3.4)      RGSC3.4
12           ggallus_gene_ensembl                 Gallus gallus genes (WASHUC2)      WASHUC2
13        tbelangeri_gene_ensembl              Tupaia belangeri genes (tupBel1)      tupBel1
14       xtropicalis_gene_ensembl             Xenopus tropicalis genes (JGI4.2)       JGI4.2
15         ecaballus_gene_ensembl                Equus caballus genes (EquCab2)      EquCab2
16          cjacchus_gene_ensembl            Callithrix jacchus genes (calJac3)      calJac3
17           pabelii_gene_ensembl                    Pongo abelii genes (PPYG2)        PPYG2
18            drerio_gene_ensembl                       Danio rerio genes (Zv9)          Zv9
19 stridecemlineatus_gene_ensembl Spermophilus tridecemlineatus genes (speTri1)      speTri1
20     tnigroviridis_gene_ensembl   Tetraodon nigroviridis genes (TETRAODON8.0) TETRAODON8.0
21        ttruncatus_gene_ensembl             Tursiops truncatus genes (turTru1)      turTru1
22       scerevisiae_gene_ensembl      Saccharomyces cerevisiae genes (SGD1.01)      SGD1.01
23      amelanoleuca_gene_ensembl        Ailuropoda melanoleuca genes (ailMel1)      ailMel1
24          celegans_gene_ensembl          Caenorhabditis elegans genes (WS220)        WS220
25          mmulatta_gene_ensembl               Macaca mulatta genes (MMUL_1.0)     MMUL_1.0
26         pvampyrus_gene_ensembl             Pteropus vampyrus genes (pteVam1)      pteVam1
27        mdomestica_gene_ensembl         Monodelphis domestica genes (monDom5)      monDom5
28            vpacos_gene_ensembl                 Vicugna pacos genes (vicPac1)      vicPac1
29     acarolinensis_gene_ensembl         Anolis carolinensis genes (AnoCar2.0)    AnoCar2.0
30         tsyrichta_gene_ensembl              Tarsius syrichta genes (tarSyr1)      tarSyr1
31        ogarnettii_gene_ensembl            Otolemur garnettii genes (otoGar1)      otoGar1
32         trubripes_gene_ensembl             Takifugu rubripes genes (FUGU4.0)      FUGU4.0
33     dmelanogaster_gene_ensembl      Drosophila melanogaster genes (BDGP5.25)     BDGP5.25
34        eeuropaeus_gene_ensembl           Erinaceus europaeus genes (eriEur1)      eriEur1
35          mmurinus_gene_ensembl            Microcebus murinus genes (micMur1)      micMur1
36          olatipes_gene_ensembl                 Oryzias latipes genes (HdrR)         HdrR
37         etelfairi_gene_ensembl              Echinops telfairi genes (TENREC)       TENREC
38     cintestinalis_gene_ensembl               Ciona intestinalis genes (JGI2)         JGI2
39      ptroglodytes_gene_ensembl             Pan troglodytes genes (CHIMP2.1)      CHIMP2.1
40         oprinceps_gene_ensembl          Ochotona princeps genes (OchPri2.0)     OchPri2.0
41          ggorilla_gene_ensembl              Gorilla gorilla genes (gorGor3)      gorGor3
42            dordii_gene_ensembl              Dipodomys ordii genes (dipOrd1)      dipOrd1
43       nleucogenys_gene_ensembl          Nomascus leucogenys genes (Nleu1.0)      Nleu1.0
44           sscrofa_gene_ensembl                   Sus scrofa genes (Sscrofa9)     Sscrofa9
45         mmusculus_gene_ensembl                 Mus musculus genes (NCBIM37)      NCBIM37
46         ocuniculus_gene_ensembl       Oryctolagus cuniculus genes (oryCun2.0)    oryCun2.0
47         mgallopavo_gene_ensembl              Meleagris gallopavo genes (UMD2)         UMD2
48           saraneus_gene_ensembl                 Sorex araneus genes (sorAra1)      sorAra1
49     dnovemcinctus_gene_ensembl          Dasypus novemcinctus genes (dasNov2)      dasNov2
50         pcapensis_gene_ensembl             Procavia capensis genes (proCap1)      proCap1
51           btaurus_gene_ensembl                   Bos taurus genes (Btau_4.0)     Btau_4.0
52          meugenii_gene_ensembl             Macropus eugenii genes (Meug_1.0)     Meug_1.0
53       cfamiliaris_gene_ensembl           Canis familiaris genes (CanFam_2.0)   CanFam_2.0
```

To select a dataset we can update the `Mart` object using the function `useDataset`. In the example below we choose to use the hsapiens dataset.

```
ensembl = useDataset("hsapiens_gene_ensembl",mart=ensembl)
```

Or alternatively if the dataset one wants to use is known in advance, we can select a BioMart database and dataset in one step by:

```
> ensembl = useMart("ensembl", dataset = "hsapiens_gene_ensembl")
```

## 3 How to build a biomaRt query

The `getBM` function has three arguments that need to be introduced: filters, attributes and values. *Filters* define a restriction on the query. For example you want to restrict the output to all genes located on the human X chromosome then the filter *chromosome_name* can be used with value 'X'. The `listFilters` function shows you all available filters in the selected dataset.

```
> filters = listFilters(ensembl)
> filters[1:5, ]
```

```
             name     description
1 chromosome_name Chromosome name
2           start Gene Start (bp)
3             end   Gene End (bp)
4      band_start      Band Start
5        band_end        Band End
```

*Attributes* define the values we are interested in to retrieve. For example we want to retrieve the gene symbols or chromosomal coordinates. The listAttributes function displays all available attributes in the selected dataset.

```
> attributes = listAttributes(ensembl)
> attributes[1:5, ]
```

```
                            name                       description
1                ensembl_gene_id                 Ensembl Gene ID
2          ensembl_transcript_id             Ensembl Transcript ID
3             ensembl_peptide_id                Ensembl Protein ID
4 canonical_transcript_stable_id Canonical transcript stable ID(s)
5                   description                       Description
```

The `getBM` function is the main query function in biomaRt. It has four main arguments:

- attributes: is a vector of attributes that one wants to retrieve (= the output of the query).

- filters: is a vector of filters that one wil use as input to the query.
- values: a vector of values for the filters. In case multple filters are in use, the values argument requires a list of values where each position in the list corresponds to the position of the filters in the filters argument (see examples below).
- mart: is and object of class `Mart`, which is created by the `useMart` function.

Note: for some frequently used queries to Ensembl, wrapper functions are available: `getGene` and `getSequence`. These functions call the `getBM` function with hard coded filter and attribute names.

Now that we selected a BioMart database and dataset, and know about attributes, filters, and the values for filters; we can build a biomaRt query. Let's make an easy query for the following problem: We have a list of Affymetrix identifiers from the u133plus2 platform and we want to retrieve the corresponding EntrezGene identifiers using the Ensembl mappings.

The u133plus2 platform will be the filter for this query and as values for this filter we use our list of Affymetrix identifiers. As output (attributes) for the query we want to retrieve the EntrezGene and u133plus2 identifiers so we get a mapping of these two identifiers as a result. The exact names that we will have to use to specify the attributes and filters can be retrieved with the `listAttributes` and `listFilters` function respectively. Let's now run the query:

```
> affyids = c("202763_at", "209310_s_at", "207500_at")
> getBM(attributes = c("affy_hg_u133_plus_2", "entrezgene"), filters = "affy_hg_u133_plus_2",
+     values = affyids, mart = ensembl)

  affy_hg_u133_plus_2 entrezgene
1         209310_s_at        837
2           207500_at        838
3           202763_at        836
```

# 4 Examples of biomaRt queries

In the sections below a variety of example queries are described. Every example is written as a task, and we have to come up with a biomaRt solution to the problem.

## 4.1 Task 1: Annotate a set of Affymetrix identifiers with HUGO symbol and chromosomal locations of corresponding genes

We have a list of Affymetrix hgu133plus2 identifiers and we would like to retrieve the HUGO gene symbols, chromosome names, start and end positions and the bands of the corresponding genes. The `listAttributes` and the `listFilters` functions give us an overview of the available attributes and filters and we look in those lists to find the corresponding attribute and filter names we need. For this query we'll need the following attributes: hgnc_symbol, chromsome_name, start_position, end_position, band and affy_hg_u133_plus_2 (as we want these in the output to provide a mapping with our original Affymetrix input identifiers. There is one filter in this query which is the affy_hg_u133_plus_2 filter as we use a list of Affymetrix identifiers as input. Putting this all together in the `getBM` and performing the query gives:

```
> affyids = c("202763_at", "209310_s_at", "207500_at")
> getBM(attributes = c("affy_hg_u133_plus_2", "hgnc_symbol", "chromosome_name", "start_position",
+     "end_position", "band"), filters = "affy_hg_u133_plus_2", values = affyids, mart = ensembl)
```

```
  affy_hg_u133_plus_2 hgnc_symbol chromosome_name start_position end_position  band
1         209310_s_at       CASP4              11      104813593    104840163 q22.3
2           207500_at       CASP5              11      104864962    104893895 q22.3
3           202763_at       CASP3               4      185548850    185570663 q35.1
```

## 4.2 Task 2: Annotate a set of EntrezGene identifiers with GO annotation

In this task we start out with a list of EntrezGene identiers and we want to retrieve GO identifiers related to biological processes that are associated with these entrezgene identifiers. Again we look at the output of `listAttributes` and `listFilters` to find the filter and attributes we need. Then we construct the following query:

```
> entrez = c("673", "837")
> goids = getBM(attributes = c("entrezgene", "go_id"), filters = "entrezgene", values = entrez,
+     mart = ensembl)
> head(goids)
```

```
   entrezgene      go_id
1         673 GO:0000186
2         673 GO:0006468
3         673 GO:0006916
4         673 GO:0007264
5         673 GO:0007268
```

### 4.3 Task 3: Retrieve all HUGO gene symbols of genes that are located on chromosomes 1,2 or Y , and are associated with one the following GO terms: "GO:0051330","GO:0000080","GO:0000114","GO:0000082" (here we'll use more than one filter)

The `getBM` function enables you to use more than one filter. In this case the filter argument should be a vector with the filter names. The values should be a list, where the first element of the list corresponds to the first filter and the second list element to the second filter and so on. The elements of this list are vectors containing the possible values for the corresponding filters.

```
go=c("GO:0051330","GO:0000080","GO:0000114"chrom=c(1,2,"Y")
getBM(attributes= "hgnc_symbol",
       filters=c("go","chromosome_name"),
       values=list(go,chrom), mart=ensembl)
```

```
  hgnc_symbol
1      PPP1CB
2       SPDYA
3       ACVR1
4        CUL3
5        RCC1
6        CDC7
7        RHOU
```

### 4.4 Task 4: Annotate set of idenfiers with INTERPRO protein domain identifiers

In this example we want to annotate the following two RefSeq identifiers: NM_005359 and NM_000546 with INTERPRO protein domain identifiers and a description of the protein domains.

```
> refseqids = c("NM_005359", "NM_000546")
> ipro = getBM(attributes = c("refseq_dna", "interpro", "interpro_description"), filt
+     values = refseqids, mart = ensembl)
```

```
ipro
  refseq_dna  interpro            interpro_description
1  NM_000546 IPR002117               p53 tumor antigen
2  NM_000546 IPR010991            p53, tetramerisation
3  NM_000546 IPR011615                p53, DNA-binding
4  NM_000546 IPR013872 p53 transactivation domain (TAD)
5  NM_000546 IPR000694              Proline-rich region
6  NM_005359 IPR001132     MAD homology 2, Dwarfin-type
7  NM_005359 IPR003619     MAD homology 1, Dwarfin-type
8  NM_005359 IPR013019               MAD homology, MH1
```

### 4.5 Task 5: Select all Affymetrix identifiers on the hgu133plus2 chip and Ensembl gene identifiers for genes located on chromosome 16 between basepair 1100000 and 1250000.

In this example we will again use multiple filters: chromosome_name, start, and end as we filter on these three conditions. Note that when a chromosome name, a start position and an end position are jointly used as filters, the BioMart webservice interprets this as return everything from the given chromosome between the given start and end positions.

```
> getBM(c("affy_hg_u133_plus_2", "ensembl_gene_id"), filters = c("chromosome_name", "start",
+     "end"), values = list(16, 1100000, 1250000), mart = ensembl)

  affy_hg_u133_plus_2 ensembl_gene_id
1           214555_at ENSG00000162009
2                     ENSG00000162009
3                     ENSG00000184471
4           205845_at ENSG00000196557
5                     ENSG00000181791
```

### 4.6 Task 6: Retrieve all entrezgene identifiers and HUGO gene symbols of genes which have a "MAP kinase activity" GO term associated with it.

The GO identifier for MAP kinase activity is GO:0004707. In our query we will use go as filter and entrezgene and hgnc_symbol as attributes. Here's the query:

```
> getBM(c("entrezgene", "hgnc_symbol"), filters = "go", values = "GO:0004707", mart = ensembl)

   entrezgene hgnc_symbol
1        5601       MAPK9
2      225689      MAPK15
3        5599       MAPK8
4        5594       MAPK1
5        6300      MAPK12
```

### 4.7 Task 7: Given a set of EntrezGene identifiers, retrieve 100bp upstream promoter sequences

All sequence related queries to Ensembl are available through the `getSequence` wrapper function. `getBM` can also be used directly to retrieve sequences but this can get complicated so using getSequence is recommended. Sequences can be retrieved using the `getSequence` function either starting from chromosomal coordinates or identifiers. The chromosome name can be specified using the *chromosome* argument. The *start* and *end* arguments are used to specify *start* and *end* positions on the chromosome. The type of sequence returned can be specified by the seqType argument

which takes the following values: 'cdna';'peptide' for protein sequences;'3utr' for 3' UTR sequences,'5utr' for 5' UTR sequences; 'gene_exon' for exon sequences only; 'transcript_exon' for transcript specific exonic sequences only;'transcript_exon_intron' gives the full unspliced transcript, that is exons + introns;'gene_exon_intron' gives the exons + introns of a gene;'coding' gives the coding sequence only;'coding_transcript_flank' gives the flanking region of the transcript including the UTRs, this must be accompanied with a given value for the upstream or downstream attribute;'coding_gene_flank' gives the flanking region of the gene including the UTRs, this must be accompanied with a given value for the upstream or downstream attribute; 'transcript_flank' gives the flanking region of the transcript exculding the UTRs, this must be accompanied with a given value for the upstream or downstream attribute; 'gene_flank' gives the flanking region of the gene excluding the UTRs, this must be accompanied with a given value for the upstream or downstream attribute.
In MySQL mode the `getSequence` function is more limited and the sequence that is returned is the 5' to 3'+ strand of the genomic sequence, given a chromosome, as start and an end position.

Task 4 requires us to retrieve 100bp upstream promoter sequences from a set of EntrzGene identifiers. The type argument in getSequence can be thought of as the filter in this query and uses the same input names given by `listFilters`. in our query we use entrezgene for the type argument. Next we have to specify which type of sequences we want to retrieve, here we are interested in the sequences of the promoter region, starting right next to the coding start of the gene. Setting the seqType to coding_gene_flank will give us what we need. The upstream argument is used to specify how many bp of upstream sequence we want to retrieve, here we'll retrieve a rather short sequence of 100bp. Putting this all together in getSequence gives:

```
> entrez = c("673", "7157", "837")
> getSequence(id = entrez, type = "entrezgene", seqType = "coding_gene_flank", upstream = 100,
+     mart = ensembl)
```

### 4.8 Task 8: Retrieve all 5' UTR sequences of all genes that are located on chromosome 3 between the positions 185514033 and 185535839

As described in the provious task getSequence can also use chromosomal coordinates to retrieve sequences of all genes that lie in the given region.

We also have to specify which type of identifier we want to retrieve together with the sequences, here we choose for entrezgene identifiers.

```
> utr5 = getSequence(chromosome = 3, start = 185514033, end = 185535839, type = "entrezgene",
+     seqType = "5utr", mart = ensembl)
> utr5
```

```
         V1               V2
  .....GAAGCGGTGGC ....   1981
```

## 4.9 Task 9: Retrieve protein sequences for a given list of EntrezGene identifiers

In this task the type argument specifies which type of identifiers we are using. To get an overview of other valid identifier types we refer to the `listFilters` function.

```
> protein = getSequence(id = c(100, 5728), type = "entrezgene", seqType = "peptide", mart = ensembl)
> protein
```

```
 peptide               entrezgene
 MAQTPAFDKPKVEL ...    100
 MTAIIKEIVSRNKRR ...   5728
```

## 4.10 Task 10: Retrieve known SNPs located on the human chromosome 8 between positions 148350 and 148612

For this example we'll first have to connect to a different BioMart database, namely snp.

```
> snpmart = useMart("snp", dataset = "hsapiens_snp")
```

The `listAttributes` and `listFilters` functions give us an overview of the available attributes and filters. From these we need: refsnp_id, allele, chrom_start and chrom_strand as attributes; and as filters we'll use: chrom_start, chrom_end and chr_name. Note that when a chromosome name, a start position and an end position are jointly used as filters, the BioMart webservice interprets this as return everything from the given chromosome between the given start and end positions. Putting our selected attributes and filters into getBM gives:

```
> getBM(c("refsnp_id", "allele", "chrom_start", "chrom_strand"), filters = c("chr_name", "chrom_start",
+     "chrom_end"), values = list(8, 148350, 148612), mart = snpmart)
```

```
   refsnp_id allele chrom_start chrom_strand
1   rs1134195    G/T      148394           -1
2   rs4046274    C/A      148394            1
3   rs4046275    A/G      148411            1
4     rs13291    C/T      148462            1
5   rs1134192    G/A      148462           -1
6   rs4046276    C/T      148462            1
7  rs12019378    T/G      148471            1
8   rs1134191    C/T      148499           -1
9   rs4046277    G/A      148499            1
10 rs11136408    G/A      148525            1
11  rs1134190    C/T      148533           -1
12  rs4046278    G/A      148533            1
13  rs1134189    G/A      148535           -1
14  rs3965587    C/T      148535            1
15  rs1134187    G/A      148539           -1
16  rs1134186    T/C      148569            1
17  rs4378731    G/A      148601            1
```

### 4.11 Task 11: Given the human gene TP53, retrieve the human chromosomal location of this gene and also retrieve the chromosomal location and RefSeq id of it's homolog in mouse.

The `getLDS` (Get Linked Dataset) function provides functionality to link 2 BioMart datasets which each other and construct a query over the two datasets. In Ensembl, linking two datasets translates to retrieving homology data across species. The usage of getLDS is very similar to `getBM`. The linked dataset is provided by a separate `Mart` object and one has to specify filters and attributes for the linked dataset. Filters can either be applied to both datasets or to one of the datasets. Use the listFilters and listAttributes functions on both `Mart` objects to find the filters and attributes for each dataset (species in Ensembl). The attributes and filters of the linked dataset can be specified with the attributesL and filtersL arguments. Entering all this information into `getLDS` gives:

```
human = useMart("ensembl", dataset = "hsapiens_gene_ensembl")
mouse = useMart("ensembl", dataset = "mmusculus_gene_ensembl")
getLDS(attributes = c("hgnc_symbol","chromosome_name", "start_position"),
       filters = "hgnc_symbol", values = "TP53",mart = human,
      attributesL = c("refseq_dna","chromosome_name","start_position"), martL = mouse)
```

```
    V1 V2      V3        V4 V5       V6
1 TP53 17 7512464 NM_011640 11 69396600
```

## 5 Using archived versions of Ensembl

It is possible to query archived versions of Ensembl through *biomaRt*. There are currently two ways to access archived versions.

## 5.1 Using the archive=TRUE

First we list the available Ensembl archives by using the `listMarts` function and setting the archive attribute to TRUE. Note that not all archives are available this way and it seems that recently this only gives access to few archives if you don't see the version of the archive you need please look at the 2nd way to access archives.

```
> listMarts(archive = TRUE)
```

```
                       biomart                    version
1              ensembl_mart_51                 Ensembl 51
2                  snp_mart_51                     SNP 51
3                 vega_mart_51                    Vega 32
4              ensembl_mart_50                 Ensembl 50
5                  snp_mart_50                     SNP 50
6                 vega_mart_50                    Vega 32
7              ensembl_mart_49   ENSEMBL GENES 49 (SANGER)
8     genomic_features_mart_49            Genomic Features
9                  snp_mart_49                        SNP
10                vega_mart_49                       Vega
11             ensembl_mart_48   ENSEMBL GENES 48 (SANGER)
12    genomic_features_mart_48            Genomic Features
13                 snp_mart_48                        SNP
14                vega_mart_48                       Vega
15             ensembl_mart_47   ENSEMBL GENES 47 (SANGER)
16    genomic_features_mart_47            Genomic Features
17                 snp_mart_47                        SNP
18                vega_mart_47                       Vega
19    compara_mart_homology_47            Compara homology
20 compara_mart_multiple_ga_47 Compara multiple alignments
21 compara_mart_pairwise_ga_47 Compara pairwise alignments
22             ensembl_mart_46   ENSEMBL GENES 46 (SANGER)
23    genomic_features_mart_46            Genomic Features
24                 snp_mart_46                        SNP
25                vega_mart_46                       Vega
26    compara_mart_homology_46            Compara homology
27 compara_mart_multiple_ga_46 Compara multiple alignments
28 compara_mart_pairwise_ga_46 Compara pairwise alignments
29             ensembl_mart_45   ENSEMBL GENES 45 (SANGER)
30                 snp_mart_45                        SNP
31                vega_mart_45                       Vega
32    compara_mart_homology_45            Compara homology
33 compara_mart_multiple_ga_45 Compara multiple alignments
34 compara_mart_pairwise_ga_45 Compara pairwise alignments
35             ensembl_mart_44   ENSEMBL GENES 44 (SANGER)
36                 snp_mart_44                        SNP
37                vega_mart_44                       Vega
38    compara_mart_homology_44            Compara homology
39 compara_mart_pairwise_ga_44 Compara pairwise alignments
40             ensembl_mart_43   ENSEMBL GENES 43 (SANGER)
41                 snp_mart_43                        SNP
42                vega_mart_43                       Vega
43    compara_mart_homology_43            Compara homology
44 compara_mart_pairwise_ga_43 Compara pairwise alignments
```

Next we select the archive we want to use using the `useMart` function, again setting the archive attribute to TRUE and giving the full name of the BioMart e.g. ensembl_mart_46.

```
> ensembl = useMart("ensembl_mart_46", dataset = "hsapiens_gene_ensembl", archive = T
```

If you don't know the dataset you want to use could first connect to the BioMart using `useMart` and then use the `listDatasets` function on this object. After you selected the BioMart database and dataset, queries can be performed in the same way as when using the current BioMart versions.

### 5.2 Accessing archives through specifying the archive host

Use the `http://www.ensembl.org` website and go down the bottom of the page. Click on 'view in Archive' and select the archive you need. Copy the url and use that url as shown below to connect to the specified BioMart database. The example below shows how to query Ensembl 54.

```
> listMarts(host = "may2009.archive.ensembl.org")
> ensembl54 = useMart(host = "may2009.archive.ensembl.org", biomart = "ENSEMBL_MART_ENSEMBL")
> ensembl54 = useMart(host = "may2009.archive.ensembl.org", biomart = "ENSEMBL_MART_ENSEMBL",
+     dataset = "hsapiens_gene_ensembl")
```

## 6 Using a BioMart other than Ensembl

To demonstrate the use of the biomaRt package with non-Ensembl databases the next query is performed using the Wormbase BioMart (WormMart). We connect to Wormbase, select the gene dataset to use and have a look at the available attributes and filters. Then we use a list of gene names as filter and retrieve associated RNAi identifiers together with a description of the RNAi phenotype.

```
> wormbase = useMart("wormbase_current", dataset = "wormbase_gene")
> listFilters(wormbase)
> listAttributes(wormbase)
> getBM(attributes = c("name", "rnai", "rnai_phenotype", "phenotype_desc"), filters = "gene_name",
+     values = c("unc-26", "his-33"), mart = wormbase)
```

| | name | rnai | rnai_phenotype | phenotype_desc |
|---|---|---|---|---|
| 1 | his-33 | WBRNAi00000104 | Emb \| Nmo | embryonic lethal \| Nuclear morphology alteration in early embryo |
| 2 | his-33 | WBRNAi00012233 | WT | wild type morphology |
| 3 | his-33 | WBRNAi00024356 | Ste | sterile |
| 4 | his-33 | WBRNAi00025036 | Emb | embryonic lethal |
| 5 | his-33 | WBRNAi00025128 | Emb | embryonic lethal |
| 6 | his-33 | WBRNAi00025393 | Emb | embryonic lethal |
| 7 | his-33 | WBRNAi00025515 | Emb \| Lva \| Unc | embryonic lethal \| larval arrest \| uncoordinated |
| 8 | his-33 | WBRNAi00025632 | Gro \| Ste | slow growth \| sterile |

```
9  his-33 WBRNAi00025686   Gro | Ste                                                 slow growth | sterile
10 his-33 WBRNAi00025785   Gro | Ste                                                 slow growth | sterile
11 his-33 WBRNAi00026259   Emb | Gro | Unc                    embryonic lethal | slow growth | uncoordinated
12 his-33 WBRNAi00026375   Emb                                                       embryonic lethal
13 his-33 WBRNAi00026376   Emb                                                       embryonic lethal
14 his-33 WBRNAi00027053   Emb | Unc                                  embryonic lethal | uncoordinated
15 his-33 WBRNAi00030041   WT                                             wild type morphology
16 his-33 WBRNAi00031078   Emb                                                       embryonic lethal
17 his-33 WBRNAi00032317   Emb                                                       embryonic lethal
18 his-33 WBRNAi00032894   Emb                                                       embryonic lethal
19 his-33 WBRNAi00033648   Emb                                                       embryonic lethal
20 his-33 WBRNAi00035430   Emb                                                       embryonic lethal
21 his-33 WBRNAi00035860   Egl | Emb                                egg laying defect | embryonic lethal
22 his-33 WBRNAi00048335   Emb | Sister Chromatid Separation abnormal (Cross-eyed)   embryonic lethal |
23 his-33 WBRNAi00049266   Emb | Sister Chromatid Separation abnormal (Cross-eyed)     embryonic lethal |
24 his-33 WBRNAi00053026   Emb | Sister Chromatid Separation abnormal (Cross-eyed)     embryonic lethal |
25 unc-26 WBRNAi00021278   WT                                                  wild type morphology
26 unc-26 WBRNAi00026915   WT                                                  wild type morphology
27 unc-26 WBRNAi00026916   WT                                                  wild type morphology
28 unc-26 WBRNAi00027544   Unc                                                        uncoordinated
29 unc-26 WBRNAi00049565   WT                                                  wild type morphology
30 unc-26 WBRNAi00049566   WT                                                  wild type morphology
```

# 7 biomaRt helper functions

This section describes a set of biomaRt helper functions that can be used to export FASTA format sequences, retrieve values for certain filters and exploring the available filters and attributes in a more systematic manner.

## 7.1 exportFASTA

The data.frames obtained by the getSequence function can be exported to FASTA files using the `exportFASTA` function. One has to specify the data.frame to export and the filename using the file argument.

## 7.2 Finding out more information on filters

### 7.2.1 filterType

Boolean filters need a value TRUE or FALSE in biomaRt. Setting the value TRUE will include all information that fulfill the filter requirement. Setting FALSE will exclude the information that fulfills the filter requirement and will return all values that don't fulfill the filter. For most of the filters, their name indicates if the type is a boolean or not and they will usually start with "with". However this is not a rule and to make sure you got the type right you can use the function `filterType` to investigate the type of the filter you want to use.

```
> filterType("with_affy_hg_u133_plus_2", ensembl)
```

```
[1] "boolean_list"
```

### 7.2.2 filterOptions

Some filters have a limited set of values that can be given to them. To know which values these are one can use the `filterOptions` function to retrieve the predetermed values of the respective filter.

```
> filterOptions("biotype", ensembl)
```

```
[1] "[IG_C_gene,IG_C_pseudogene,IG_D_gene,IG_J_gene,IG_J_pseudogene,IG_V_gene,IG_V_pseudogene
```

If there are no predetermed values e.g. for the entrezgene filter, then `filterOptions` will return the type of filter it is. And most of the times the filter name or it's description will suggest what values one case use for the respective filter (e.g. entrezgene filter will work with enterzgene identifiers as values)

## 7.3 Attribute Pages

For large BioMart databases such as Ensembl, the number of attributes displayed by the `listAttributes` function can be very large. In BioMart databases, attributes are put together in pages, such as sequences, features, homologs for Ensembl. An overview of the attributes pages present in the respective BioMart dataset can be obtained with the `attributePages` function.

```
> pages = attributePages(ensembl)
> pages
```

```
[1] "feature_page"     "structure"        "transcript_event" "homologs"         "snp"
```

To show us a smaller list of attributes which belog to a specific page, we can now specify this in the `listAttributes` function as follows:

```
> listAttributes(ensembl, page = "feature_page")
```

```
                                        name                                    des
1                            ensembl_gene_id                            Ensembl
2                      ensembl_transcript_id                          Ensembl Trans
3                         ensembl_peptide_id                             Ensembl Pr
4           canonical_transcript_stable_id               Canonical transcript stab
5                                description                                   Des
```

| | | |
|---|---|---|
| 6 | chromosome_name | Chromos |
| 7 | start_position | Gene St |
| 8 | end_position | Gene |
| 9 | strand | |
| 10 | band | |
| 11 | transcript_start | Transcript St |
| 12 | transcript_end | Transcript |
| 13 | external_gene_id | Associated G |
| 14 | external_transcript_id | Associated Transcr |
| 15 | external_gene_db | Associated |
| 16 | transcript_db_name | Associated Trans |
| 17 | transcript_count | Transcri |
| 18 | percentage_gc_content | % GC |
| 19 | gene_biotype | Gene |
| 20 | transcript_biotype | Transcript |
| 21 | source | |
| 22 | status | Statu |
| 23 | transcript_status | Status (tra |
| 24 | go_id | GO Term Access |
| 25 | name_1006 | GO Term N |
| 26 | definition_1006 | GO Term Definit |
| 27 | go_linkage_type | GO Term Evidence C |
| 28 | namespace_1003 | G |
| 29 | goslim_goa_accession | GOSlim GOA Acce |
| 30 | goslim_goa_description | GOSlim GOA Des |
| 31 | ox_pubmed__dm_dbprimary_acc_1074 | P |
| 32 | ucsc | |
| 33 | pdb | |
| 34 | clone_based_ensembl_gene_name | Clone based Ensembl g |
| 35 | clone_based_ensembl_transcript_name | Clone based Ensembl transcr |
| 36 | clone_based_vega_gene_name | Clone based VEGA g |
| 37 | clone_based_vega_transcript_name | Clone based VEGA transcr |
| 38 | ccds | |
| 39 | embl | EMBL (Gen |
| 40 | ens_hs_gene | Ensembl to LRG link |
| 41 | ens_hs_transcript | Ensembl to LRG link transc |
| 42 | ens_hs_translation | Ensembl to LRG link transla |
| 43 | ox_ens_lrg_transcript__dm_dbprimary_acc_1074 | LRG to Ensembl link tr |
| 44 | entrezgene | Entre |
| 45 | ottt | VEGA transcript ID(s |
| 46 | ottg | VEGA gene ID(s |
| 47 | shares_cds_with_enst | Ensembl transcript (where OTTT shares CDS wi |
| 48 | shares_cds_with_ottt | HAVANA transcript (where ENST shares CDS wi |
| 49 | shares_cds_and_utr_with_ottt | HAVANA transcript (where ENST identical |
| 50 | hgnc_id | HG |

| | | |
|---|---|---|
| 51 | hgnc_symbol | HGN |
| 52 | hgnc_transcript_name | HGNC transcr |
| 53 | ipi | |
| 54 | merops | M |
| 55 | mim_morbid_accession | MIM Morbid A |
| 56 | mim_morbid_description | MIM Morbid Des |
| 57 | mim_gene_accession | MIM Gene A |
| 58 | mim_gene_description | MIM Gene Des |
| 59 | mirbase_accession | miRBase Acce |
| 60 | mirbase_id | miRBa |
| 61 | mirbase_gene_name | miRBase g |
| 62 | mirbase_transcript_name | miRBase transcr |
| 63 | protein_id | Protein (Gen |
| 64 | refseq_dna | RefSe |
| 65 | refseq_dna_predicted | RefSeq Predicte |
| 66 | refseq_rna_predicted | RefSeq RNA p |
| 67 | refseq_peptide | RefSeq Pr |
| 68 | refseq_peptide_predicted | RefSeq Predicted Pr |
| 69 | refseq_genomic | RefSeq Genom |
| 70 | rfam | |
| 71 | rfam_gene_name | Rfam g |
| 72 | rfam_transcript_name | Rfam transcr |
| 73 | unigene | Un |
| 74 | uniprot_sptrembl | UniProt/TrEMBL A |
| 75 | uniprot_swissprot | UniProt/Swis |
| 76 | uniprot_swissprot_accession | UniProt/SwissProt A |
| 77 | uniprot_genename | UniProt G |
| 78 | wikigene_name | WikiG |
| 79 | wikigene_description | WikiGene des |
| 80 | hpa | Human Protein Atlas Ant |
| 81 | dbass3_id | Database of Aberrant 3' Splice Sites (DBA |
| 82 | dbass3_name | DBASS3 G |
| 83 | dbass5_id | Database of Aberrant 5' Splice Sites (DBA |
| 84 | dbass5_name | DBASS5 G |
| 85 | affy_hc_g110 | Affy |
| 86 | affy_hg_focus | Affy |
| 87 | affy_hg_u133_plus_2 | Affy HG U13 |
| 88 | affy_hg_u133a_2 | Affy HG |
| 89 | affy_hg_u133a | Affy |
| 90 | affy_hg_u133b | Affy |
| 91 | affy_hg_u95av2 | Affy H |
| 92 | affy_hg_u95b | Affy |
| 93 | affy_hg_u95c | Affy |
| 94 | affy_hg_u95d | Affy |
| 95 | affy_hg_u95e | Affy |

```
96                                affy_hg_u95a                                Affy
97                                affy_hugenefl                               Affy H
98                              affy_huex_1_0_st_v2                           Affy HuEx 1
99                             affy_hugene_1_0_st_v1                         Affy HuGene 1
100                                affy_u133_x3p                                Affy
101                              agilent_cgh_44b                             Agilent
102                            agilent_wholegenome                           Agilent Who
103                                       codelink                                  
104                           illumina_humanwg_6_v1                         Illumina Huma
105                           illumina_humanwg_6_v2                         Illumina Huma
106                           illumina_humanwg_6_v3                         Illumina Huma
107                             illumina_humanht_12                          Illumina Hum
108                                phalanx_onearray                              Phalanx
109                              anatomical_system                   Anatomical System (eg
110                              development_stage                   Development Stage (eg
111                                       cell_type                          Cell Type (eg
112                                      pathology                           Pathology (eg
113                                  atlas_celltype                            GNF/Atlas c
114                               atlas_diseasestate                         GNF/Atlas disea
115                               atlas_organismpart                         GNF/Atlas organ
116                               family_description                      Ensembl Family Des
117                                          family                     Ensembl Protein Fami
118                                           pirsf                          PIRSF SuperF
119                                     superfamily                               Superf
120                                           smart                                  
121                                         profile                                PR
122                                          prints                                 P
123                                            pfam                                 
124                                         tigrfam                                TI
125       protein_feature_seg__dm_hit_name_1048                                    
126                                        interpro                                Int
127                      interpro_short_description                   Interpro Short Des
128                            interpro_description                         Interpro Des
129                            transmembrane_domain                         Transmembran
130                                  signal_domain                                Signa
131                                          ncoils                                  
```

We now get a short list of attributes related to the region where the genes are located.

# 8 Local BioMart databases

The biomaRt package can be used with a local install of a public BioMart database or a locally developed BioMart database and web service. In order

for biomaRt to recognize the database as a BioMart, make sure that the local database you create has a name conform with

```
database_mart_version
```

where database is the name of the database and version is a version number. No more underscores than the ones showed should be present in this name. A possible name is for example

```
ensemblLocal_mart_46
```

.

### 8.1 Minimum requirements for local database installation

More information on installing a local copy of a BioMart database or develop your own BioMart database and webservice can be found on `http://www.biomart.org` Once the local database is installed you can use biomaRt on this database by:

```
listMarts(host="www.myLocalHost.org", path="/myPathToWebservice/martservice")
mart=useMart("nameOfMyMart",dataset="nameOfMyDataset",host="www.myLocalHost.org", path="/myPathToWebservice/martser
```

For more information on how to install a public BioMart database see: http://www.biomart.org/install.html and follow link databases.

## 9 Session Info

```
> sessionInfo()

R version 2.13.0 (2011-04-13)
Platform: i386-pc-mingw32/i386 (32-bit)

locale:
[1] LC_COLLATE=C                          LC_CTYPE=English_United States.1252    LC_
[4] LC_NUMERIC=C                          LC_TIME=English_United States.1252

attached base packages:
[1] stats     graphics  grDevices utils     datasets  methods   base

other attached packages:
[1] biomaRt_2.8.1

loaded via a namespace (and not attached):
[1] RCurl_1.6-5.1 XML_3.4-0.2   tools_2.13.0
```

```
> warnings()

NULL
```