

# Estimating the Number of Essential Genes using Occugene

Oliver Will

November 6, 2019

This vignette contains code from a chapter of the forthcoming book, A Osterman and S Gerdes. *Gene Essentiality: Protocols and Bioinformatics*. Humana Press. This package has similar functionality as the R package **negenes** written by Karl Broman.

Biologists have built large random mutagenesis libraries for prokaryotes. For a description of the biology, see MA Jacobs (et al). (2003) Comprehensive transposon mutant library of *Pseudomonas aeruginosa*. *Proc Natl Acad Sci USA*. **100**:14339–14344. The **occugene** package provides statistical tools to help build random libraries.

We model the number of insertions per clone as a Multinomial( $n, p_1, \dots, p_k$ ) random vector. The number of knockouts in the library follows the occupancy distribution of the multinomial random variable. We compute the expected number of genes hit if there were no essential genes.

```
> library("occugene")
> n <- 60
> p <- c(seq(10, 1, -1), seq(10, 1, -1), 18)/124
> p <- p/sum(p)
> eMult(n, p)
```

```
[1] 17.74773
```

```
> varMult(n, p)
```

```
[1] 1.744004
```

We approximate the moments of the occupancy distribution using Monte Carlo integration.

```
> eMult(n, p, iter=100, seed=4)
```

```
[1] 17.64
```

```
> varMult(n, p, iter=100, seed=4)
```

```
[1] 2.050909
```

We load an example hit table and experimental results to analyze. The format of the hit table is different than what is used in **negenes** because we wish to track the order of insert locations.

```
> data(sampleAnnotation)
> data(sampleInsertions)
> print(sampleAnnotation)
```

	idNum	first	last	orientation
1	1	2	11	0
2	2	13	21	0
3	3	23	30	0
4	4	32	38	0
5	5	40	45	0
6	6	44	48	0
7	7	50	53	0
8	8	55	57	0
9	9	59	60	0
10	10	62	62	0
11	11	64	73	0
12	12	75	83	0
13	13	85	92	0
14	14	94	100	0
15	15	102	107	0
16	16	106	110	0
17	17	112	115	0
18	18	117	119	0
19	19	121	122	0
20	20	124	124	0

```
> print(sampleInsertions)
```

	position
1	69
2	2
3	36
4	34
5	99
6	33
7	91
8	113
9	120
10	8
11	93
12	35

13	11
14	120
15	52
16	55
17	122
18	69
19	121
20	94
21	90
22	124
23	62
24	61
25	84
26	100
27	58
28	101
29	63
30	64
31	68
32	31
33	111
34	84
35	58
36	122
37	56
38	72
39	49
40	2
41	116
42	31
43	67
44	18
45	113
46	9
47	113
48	112
49	91
50	67
51	49
52	93
53	112
54	98
55	99
56	52
57	17
58	17

```
59      112
60      92
```

```
> a.data <- sampleAnnotation
> experiment <- sampleInsertions
```

We estimate the number of genes that will be knocked out in the next 10 clones using the Efron and Thisted estimator.

```
> orf <- cbind(a.data$first,a.data$last)
> clone <- experiment$position
> etDelta(10,orf,clone)
```

```
$expected
[1] 0.1190665
```

```
$variance
[1] 0.02936508
```

We use the Will and Jacobs' bootstrap to estimate the number of knockouts made in the next 10 clones.

```
> orf <- cbind(a.data$first,a.data$last)
> clone <- experiment$position
> fFit(orf,clone,FALSE)
```

```
Nonlinear regression model
model: noOrfs ~ b0 - b1 * exp(-b2 * x)
data: cumul
      b0      b1      b2
12.34393 12.05475 0.06668
residual sum-of-squares: 15.62
```

```
Number of iterations to convergence: 6
Achieved convergence tolerance: 7.669e-07
```

```
> unbiasedDelta0(10,orf,clone,iter=10,seed=4,alpha=0.05,TR=F)
```

```
$delta0
[1] 0.2959099
```

```
$CI
[1] -0.1730529 0.5010068
```

We estimate the number of essential genes using the Will and Jacobs' bootstrap.

```
> unbiasedB0(orf,clone,iter=10,seed=4,alpha=0.05,TR=F)
```

```
$b0  
[1] 14.42681
```

```
$CI  
[1] 10.87780 16.83998
```

Finally, we convert `occugene`'s data format into the format for `negenes`.

```
> newOrf <- occup2Negenes(orf,clone)  
> print(newOrf)
```

	n.sites	n.sites2	counts	counts2
1	10	0	5	0
2	9	0	3	0
3	8	0	0	0
4	7	0	4	0
5	4	2	0	0
6	3	0	0	0
7	4	0	2	0
8	3	0	2	0
9	2	0	0	0
10	1	0	1	0
11	10	0	7	0
12	9	0	0	0
13	8	0	4	0
14	7	0	5	0
15	4	2	0	0
16	3	0	0	0
17	4	0	6	0
18	3	0	0	0
19	2	0	3	0
20	1	0	1	0

We outlined the basic features of the `occugene` package in this Sweave document.