

An Introduction to *PROMISE*

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1 Introduction

PROMISE, PROjection onto the Most Interesting Statistical Evidence, is a general procedure to identify genomic features that exhibit a specific biologically interesting pattern of association with multiple phenotypic endpoint variables. Biological knowledge of the endpoint variables is used to define a vector that represents the biologically most interesting values for a set of association statistics. If prior biological knowledge of the endpoint variables is not available, a projection to point 0 is performed. The PROMISE performs one hypothesis test for each genomic feature, and is flexible to accommodate various types of endpoints. In this update, we also incorporate a fast permutation based on negative binomial sampling strategy, in which further permutation for a gene will not be performed for a gene with fixed number of success achieved.

In this document, we describe how to perform PROMISE procedure using hypothetical example data sets provided with the package.

2 Requirements

The PROMISE package depends on *Biobase* and *GSEABase*. The understanding of *ExpressionSet* and *GeneSetCollection* is a prerequisite to perform the PROMISE procedure. Due to the internal handling of multiple endpoints, the consistency of *ExpressionSet* and *GeneSetCollection* is assumed. The detailed requirements are illustrated below.

Load the PROMISE package and the example data sets: `sampExprSet`, `sampGeneSet`, and `phPatt` into R.

```
> library(PROMISE)
> data(sampExprSet)
> data(sampGeneSet)
> data(phPatt)
```

The *ExpressionSet* should contain at least two components: *exprs* (array data) and *phenoData* (endpoint data). *exprs* is a data frame with column names representing the array identifiers (IDs) and row names representing the

probe (genomic feature) IDs. *phenoData* is an *AnnotatedDataFrame* with column names representing the endpoint variables and row names representing array. The array IDs of *phenoData* and *exprs* should be matched.

```
> arrayData<-exprs(sampExprSet)
> ptData<- pData(phenoData(sampExprSet))
> head(arrayData[, 1:4])
```

	array_1	array_2	array_3	array_4
probe_1	7.594327	6.710827	8.151585	5.885169
probe_2	4.719728	5.611497	5.819049	3.896193
probe_3	8.815163	5.451243	6.670862	7.295233
probe_4	5.187320	5.142358	6.105922	5.333343
probe_5	4.102526	2.655307	4.139362	1.801595
probe_6	5.415526	4.498429	4.393805	5.219712

```
> head(ptData)
```

	drugLevel	residualDisease	obsTime	obsCensor	strat
array_1	2.486889		3 0.6117827	1	2
array_2	2.931522		3 1.2657796	1	2
array_3	2.569262		3 1.6318570	0	2
array_4	2.157207		2 0.3879765	0	1
array_5	1.649005		3 0.5763599	1	2
array_6	2.795997		3 0.5474852	0	1

```
> all(colnames(arrayData)==rownames(ptData))
```

```
[1] TRUE
```

The *GeneSetCollection* should be extractable in the following way. The probe IDs should be a subset of probe IDs of *exprs*.

```
> GS.data<-NULL
> for (i in 1:length(sampGeneSet)){
+   tt<-sampGeneSet[i][[1]]
+   this.name<-unlist(geneIds(tt))
+   this.set<-setName(tt)
+   this.data<- cbind.data.frame(featureID=as.character(this.name),
+                                 setID=rep(as.character(this.set), length(this.name)))
+   GS.data<-rbind.data.frame(GS.data, this.data)
+ }
> sum(!is.element(GS.data[,1], rownames(arrayData)))==0
```

```
[1] TRUE
```

The association pattern definition is critical. The prior biological knowledge is required to define the vector that represents the biologically most interesting

values for statistics. If prior biological knowledge of the endpoint variables cannot be assumed, an arbitrary *stat.coef* can be used as it will be ignored with *proj0=TRUE*. In this hypothetical example, we are interested in identifying genomic features that are positively associated with active drug level, negatively associated with minimum disease, and positively associated with survival. The three endpoints are represented in three rows as shown below:

```
> phPatt
```

	stat.coef	stat.func	endpt.vars
Association 1	1	spearman.rstat	drugLevel
Association 2	-1	spearman.rstat	residualDisease
Association 3	1	jung.rstat	obsTime,obsCensor

3 PROMISE Analysis

As mentioned in section 2, the *ExpressionSet* and pattern definition are required by PROMISE procedure. *GeneSetCollection* is required if a gene set enrichment analysis (GSEA) is to be performed within the PROMISE analysis.

The code below performs a PROMISE analysis without GSEA. As mentioned above, *GeneSetCollection* is not needed. The gene set result is NULL.

```
> test1 <- PROMISE(exprSet=sampExprSet,
+                  geneSet=NULL,
+                  promise.pattern=phPatt,
+                  strat.var=NULL,
+                  proj0=FALSE,
+                  nbperm=FALSE,
+                  max.ntail=10,
+                  seed=13,
+                  nperms=100)
```

Gene level (genomic feature) result:

```
> gene.res<-test1$generes
> head(gene.res)
```

	probeid	drugLevel.stat	residualDisease.stat	
1	probe_1	0.50745310	-0.4410378	
2	probe_2	-0.05129198	0.1083797	
3	probe_3	0.34392723	-0.3069893	
4	probe_4	-0.21510307	0.1812378	
5	probe_5	-0.43215514	0.3725875	
6	probe_6	0.06213032	-0.1796821	
	obsTime.obsCensor.stat	PROMISE.stat	drugLevel.perm.p	
1	-0.328713893	0.20659234	0.00	
2	0.050025633	-0.03654867	0.72	

3	-0.277173083	0.12458114	0.00
4	0.003140076	-0.13106692	0.19
5	0.346205081	-0.15284585	0.01
6	-0.201508221	0.01343473	0.63
	residualDisease.perm.p	obsTime.obsCensor.perm.p	
1	0.00	0.00	
2	0.42	0.64	
3	0.02	0.07	
4	0.21	0.99	
5	0.03	0.00	
6	0.14	0.18	
	PROMISE.perm.p		
1	0.00		
2	0.65		
3	0.08		
4	0.05		
5	0.03		
6	0.86		

Gene set level result:

```
> set.res<-test1$setres
> head(set.res)
```

NULL

The code below performs a PROMISE analysis with GSEA and using fast permutation. As mentioned above, *GeneSetCollection* is required. *sampGeneSet*, a *GeneSetCollection*, is passed as an argument to PROMISE.

```
> test2 <- PROMISE(exprSet=sampExprSet,
+                  geneSet=sampGeneSet,
+                  promise.pattern=phPatt,
+                  strat.var=NULL,
+                  proj0=FALSE,
+                  nbperm=TRUE,
+                  max.ntail=10,
+                  seed=13,
+                  nperms=100)
```

Gene level (genomic feature) result:

```
> gene.res2<-test2$generes
> head(gene.res2)
```

	probeid	drugLevel.stat	residualDisease.stat
1	probe_1	0.50745310	-0.4410378
2	probe_2	-0.05129198	0.1083797

3	probe_3	0.34392723	-0.3069893
4	probe_4	-0.21510307	0.1812378
5	probe_5	-0.43215514	0.3725875
6	probe_6	0.06213032	-0.1796821

	obsTime.obsCensor.stat	PROMISE.stat	drugLevel.perm.p
1	-0.328713893	0.20659234	0.00
2	0.050025633	-0.03654867	0.72
3	-0.277173083	0.12458114	0.00
4	0.003140076	-0.13106692	0.19
5	0.346205081	-0.15284585	0.01
6	-0.201508221	0.01343473	0.63

	residualDisease.perm.p	obsTime.obsCensor.perm.p
1	0.00	0.00
2	0.42	0.64
3	0.02	0.07
4	0.21	0.99
5	0.03	0.00
6	0.14	0.18

	PROMISE.perm.p	nperms
1	0.00	100
2	0.65	100
3	0.08	100
4	0.05	100
5	0.03	100
6	0.86	100

Gene set level result:

```
> set.res2<-test2$setres
> head(set.res2)
```

	setid	drugLevel.stat	residualDisease.stat
1	GeneSet 1	0.30998611	0.28204640
2	GeneSet 2	0.18012248	0.15382306
3	GeneSet 3	0.19852231	0.18092662
4	GeneSet 4	0.07289261	0.13631292
5	GeneSet 5	0.11745341	0.06539079
6	GeneSet 6	0.08167737	0.08478505

	obsTime.obsCensor.stat	PROMISE.stat	drugLevel.perm.p
1	0.20105155	0.13032699	0.0000000
2	0.13759326	0.09124962	0.0000000
3	0.20620859	0.06608593	0.0600000
4	0.12875576	0.04299261	0.6888889
5	0.09114311	0.06327372	0.4700000
6	0.09922607	0.03234743	0.6956522

	residualDisease.perm.p	obsTime.obsCensor.perm.p
1	0.0000000	0.0200000

2	0.0200000	0.1500000
3	0.0900000	0.0600000
4	0.3111111	0.3111111
5	0.8700000	0.7100000
6	0.6521739	0.5217391

	PROMISE.perm.p	nperms
1	0.0000000	100
2	0.0000000	100
3	0.3200000	100
4	0.7888889	90
5	0.2500000	100
6	0.9565217	23

The code below performs a PROMISE analysis with GSEA and using fast permutation without prior knowledge of the three endpoint variables (*proj0=TRUE*). As mentioned above, *GeneSetCollection* is required. *sampGeneSet*, a *GeneSetCollection*, is passed as an argument to PROMISE.

```
> test3 <- PROMISE(exprSet=sampExprSet,
+                  geneSet=sampGeneSet,
+                  promise.pattern=phPatt,
+                  strat.var=NULL,
+                  proj0=TRUE,
+                  nbperm=TRUE,
+                  max.ntail=10,
+                  seed=13,
+                  nperms=100)
```

Gene level (genomic feature) result:

```
> gene.res3<-test3$generes
> head(gene.res3)
```

	probeid	drugLevel.stat	residualDisease.stat
1	probe_1	0.50745310	-0.4410378
2	probe_2	-0.05129198	0.1083797
3	probe_3	0.34392723	-0.3069893
4	probe_4	-0.21510307	0.1812378
5	probe_5	-0.43215514	0.3725875
6	probe_6	0.06213032	-0.1796821

	obsTime.obsCensor.stat	PROMISE.stat	drugLevel.perm.p
1	-0.328713893	0.56007583	0.00
2	0.050025633	0.01687958	0.72
3	-0.277173083	0.28935328	0.00
4	0.003140076	0.07912632	0.19
5	0.346205081	0.44543747	0.01
6	-0.201508221	0.07675139	0.63

	residualDisease.perm.p	obsTime.obsCensor.perm.p
1	0.00	0.00
2	0.42	0.64
3	0.02	0.07
4	0.21	0.99
5	0.03	0.00
6	0.14	0.18

	PROMISE.perm.p	nperms
1	0.00	100
2	0.74	100
3	0.02	100
4	0.30	100
5	0.00	100
6	0.21	100

Gene set level result:

```
> set.res3<-test3$setres
> head(set.res3)
```

	setid	drugLevel.stat	residualDisease.stat
1	GeneSet 1	0.30998611	0.28204640
2	GeneSet 2	0.18012248	0.15382306
3	GeneSet 3	0.19852231	0.18092662
4	GeneSet 4	0.07289261	0.13631292
5	GeneSet 5	0.11745341	0.06539079
6	GeneSet 6	0.08167737	0.08478505

	obsTime.obsCensor.stat	PROMISE.stat	drugLevel.perm.p
1	0.20105155	0.27817450	0.0000000
2	0.13759326	0.15129808	0.0000000
3	0.20620859	0.16602416	0.0000000
4	0.12875576	0.06543595	0.8400000
5	0.09114311	0.04097703	0.3400000
6	0.09922607	0.03950889	0.6229508

	residualDisease.perm.p	obsTime.obsCensor.perm.p
1	0.0000000	0.0200000
2	0.0200000	0.1500000
3	0.0700000	0.0100000
4	0.3000000	0.2900000
5	0.7100000	0.5100000
6	0.6393443	0.4918033

	PROMISE.perm.p	nperms
1	0.0000000	100
2	0.0000000	100
3	0.0000000	100
4	0.3400000	100

5	0.8300000	100
6	0.8196721	61